|--|

Award Number: DAMD17-98-1-8150

TITLE: Extracellular Matrix in Breast Cancer Invasion

PRINCIPAL INVESTIGATOR:

Vito Quaranta, M.D.

CONTRACTING ORGANIZATION:

The Scripps Research Institute

La Jolla, California 92037

REPORT DATE:

June 2000

TYPE OF REPORT:

Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release; Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE			Form Approved OMB No. 074-0188	
Public reporting burden for this collection of inform the data needed, and completing and reviewing th reducing this burden to Washington Headquarters Management and Budget, Paperwork Reduction P	ation is estimated to average 1 hour per response, is collection of information. Send comments regard	including the time for reviewing ins ing this burden estimate or any oth	tructions, searching ex er aspect of this collec	sting data sources, gathering and maintaining tion of information, including suggestions for
reducing this burden to Washington Headquarters Management and Budget, Paperwork Reduction P	Services, Directorate for Information Operations an roject (0704-0188), Washington, DC 20503	d Reports, 1215 Jefferson Davis H	Inghway, Suite 1204, Ar	ington, VA ZZZUZ-43UZ, and to the Office of
1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE June 2000	3. REPORT TYPE AND	- 31 May 00)	: U
4. TITLE AND SUBTITLE			5. FUNDING N	:
"Extracellular Mate	cix in Breast Cancer	r Invasion"	DWIDT 1-38.	-1-0130
6. AUTHOR(S)				
Vito Quaranta, M.D.				
7. PERFORMING ORGANIZATION N	AME(S) AND ADDRESS(ES)			G ORGANIZATION
The Scripps Research Institute La Jolla, California 92037			REPORT NU	MBER
E-Mail: quaranta@scrlpps.edu		·		
9. SPONSORING / MONITORING A	GENCY NAME(S) AND ADDRESS(ES)		NG / MONITORING EPORT NUMBER
U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			AGENOTA	EI GIT HOMBER
AA OUDDI Primita birina				
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited				12b. DISTRIBUTION CODE
metastasis as a breakder that metastasis is initial motility. Therefore, or interfering with, molecthat, in the mammary grant matrix metalloproteases the breast gland basem identification of the position, relative to motility. A second imposition information is critical to the ability of antibod hopefully metastasis.	t is to find novel treatown of mechanisms that called by molecular cuerur approach is to block cular and cellular mechaland, several of these is (MMP) with laminin-5 ent membrane. One major Ln-5 site onto which cethis site, of the docking portant finding is the on of the Ln-5 fragment itical to design in vivies to Ln-5, or Ln-5 fragment.	govern tissue or s that improperl metastasis by userisms that regumechanisms revolute. (Ln-5), an extrair finding in thills adhere and mand site for antillocation of Ln-5 resulting from a animal experimes.	ganization y stimulat understandi ulate cell ve around ucellular m is past yea nigrate. W bodies tha is sites tha uthis prot ments in wh	. We postulated e cancer cell ng, and then motility. We found the interaction of atrix protein of r was the e also defined the t block cell t are cleaved by eolytic activity. ich we will test ll motility and
14. SUBJECT TERMS				15. NUMBER OF PAGES
Metastasis Cell Migration Laminin-5 Metalloproteases				16. PRICE CODE
17. SECURITY CLASSIFICATION	18. SECURITY CLASSIFICATION	19. SECURITY CLASSIF	FICATION	20. LIMITATION OF ABSTRACT
OF REPORT Unclassified	OF THIS PAGE Unclassified	OF ABSTRACT Unclassif	ied	Unlimited

Unclassified

Unclassified

Unlimited

FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

Where copyrighted material is quoted, permission has been obtained to use such material.

Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

N/A In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and use of Laboratory Animals of the Institute of Laboratory Resources, national Research Council (NIH Publication No. 86-23, Revised 1985).

For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.

PI - Signature

Table of Contents

Front Cover	1
SF298	2
Foreword	3
Table of Contents	4
Introduction	5
Body	5
Key Research Accomplishments	6
Reportable Outcomes	6
Conclusions	7
References	7
Appendices	7

Progress Report 1999-2000 (REVISED)
Idea Grant DAMD 17-98-1-8150
P.I. Vito Quaranta

Introduction (paraphrased from last year's):

Our long-term goal is to find new treatments for breast cancer metastasis, based on targeting cellular and molecular mechanisms underlying the process of metastasis itself. The element of innovation in our approach is that we visualize metastasis as a problem of breakdown in tissue organization. Consequently, for the purposes of cancer treatment, our target is the mammary gland as a tissue, rather than the individual cells. By taking a reductionist approach, we investigate the actual molecular mechanisms that keep breast epithelial cells segregated on the luminal side of the basal lamina. These cells are the ones from which invasive breast cancer arises. We previously identified a molecular mechanism (1, 2) that determines whether normal or cancer breast cells may cross the basal lamina. This mechanism relies on the interaction of laminin-5, a major extracellular matrix molecule of basal lamina, with matrix metalloproteases and integrins. The challenge of this proposal is to determine how dominant this mechanism is in regulating migratory versus stationary behavior of breast epithelial cells. Should we find that such mechanism is rate limiting for metastasis, we will be close to a discovery phase for novel drugs or treatments that may prevent or block breast cancer invasion.

Body:

AIM 1. To inhibit mammary epithelial cell motility in vitro and cell metastasis in vivo by blocking the migratory site of laminin-5 (Ln-5).

MIG-1 is an antibody that blocks migration of cancer cells on Ln-5 cleaved by metalloproteases (MMPs). We proposed to map the binding site of this antibody on the Ln-5 molecule, since it might unveil structural features of Ln-5 that stimulate migration. We have now completed the mapping of the MIG-1 epitope on the LG2 domain of the Ln-5 α 3 subunit. By site-directed mutagenesis, we showed that the epitope is restricted to a stretch of 6 amino acids towards the center of the molecule. Recently, the three-dimensional structure of the Ln-2 α 2LG5 domain was solved. This domain is structurally homologous to α 3LG2. Therefore, we were able to model the primary sequence of LG2 over the structure of LG5. This comparison predicted that the MIG-1 epitope would be located in a loop of the LG5 domain which is exposed to the aqueous environment, corroborating our mapping results.

To prove that the LG2 domain is involved in cell adhesion and migration, we also mapped the integrin binding site of Ln-5. Unexpectedly, the integrin binding site maps not to LG2, but rather to the LG3 domain, which is spatially close to LG2. This result suggest a mechanism whereby the LG2 and LG3 domain interact with each other and with integrins in order to support adhesion and migration. This mechanism is currently being investigated.

AIM 2. To inhibit mammary epithelial cell motility in vitro and cell metastasis in vivo by inhibiting the cleavage of Ln-5 by MMP2.

During last year, we found that MMP2 is not the only protease to cleave Ln-5. MT1-MMP, a surface bound MMP, cleaves Ln-5 at the same site as MMP2, as well as to another site, approximately 150 amino acids upstream (1). The interesting fact is that the combined actions of MMP2 and MT1-MMP liberate a fragment of the Ln-5 γ 2 chain, whose structure resembles that of EGF (epidermal growth factor). EGF and EGF-like ligands are well known to display mitogenic and motogenic activity. We have made recombinant proteins spanning the cleavage sites of these MMPs, as well as the liberated fragment. By using these recombinant fragments with appropriate tags, we have confirmed that both MMP2 and MT1-MMP cleave them in vitro. Furthermore, some of these fragments interfered with cell migration in in vitro assays. An intriguing result is that the liberated fragment binds to the cell surface and stimulated tyrosine phosphorylation, suggesting it may bind to a receptor. These results strongly encourage testing these recombinant fragments in vivo, which we plan to carry out this coming year.

Aim 3. To produce monoclonal antibodies that react with MMP2-cleaved Ln-5 and not with intact Ln-5, and to use them in immunohistological assays for correlating the location of cleaved Ln-5 with breast cancer cell invasion sites.

Using purified Ln-5 for this purpose has proven not feasible, because both intact and cleaved Ln-5 are present in purified preparations and are difficult to separate. We plan to change approach and use instead the recombinant fragments mentioned in the Aim above.

Key Research Accomplishments:

- Mapped integrin binding site on Laminin-5 to domain LG3, first time this was done on any laminins
- Mapped epitope for antibody MIG-1, which blocks cell migration, to Laminin-5 domain LG2
- Discovered that MT1-MMP is another proteolytic enzyme, besides MMP2, which cleaves Laminin-5
- Defined the boundaries of a Laminin-5 fragment that is proteolytically cleaved out by the action of MMPs
- Produced recombinant Laminin-5 MMP fragments

Reportable Outcomes:

- Manuscripts, abstracts, presentations

Two manuscripts in print, see References below

Presentation: 2000 Gordon Conference "Basement Membranes"

Patents and licenses

None

- Degrees obtained

None

- Development of reagents

Recombinant Domain III from Laminin-5
Recombinant LG2 and LG3 domains from Laminin-5

- Informatics

None

- Funding applied for

NIH grant application RO1-GM46902 "Molecular Regulation of Integrin Functions by Laminin Domains", PI Vito Quaranta, based on results from Aims 1 and 2 of this grant.

- Employment

None

Conclusions:

We mapped the Ln-5 binding site for antibodies that block adhesion and/or migration. In addition, we identified the domain of Ln-5 that binds integrins and supports cell adhesion and migration. Surprisingly, this domain is close to, but does not overlap with the binding site of the blocking antibodies. Nonetheless, this is an important insight towards our goal of designing reagents that inhibit the migratory site of Ln-5 and therefore may interfere with metastasis.

We have overcome limitations imposed by protein purification methods, and plan to produce cleaved Ln-5 monoclonal antibodies by using recombinant fragments as immunogens.

We have further defined Ln-5 fragments that are the targets of, or result from MMP cleavage, and are closer to test them in vivo for possible inhibitory effects on metastasis. This has been a long-standing question in the Ln-5 field, and therefore we expect our results to have significant impact. In spite of the fact that Ln-5 is clearly involved in metastasis, absence of structural details on its adhesion/migration domains has frustrated efforts to interfere with cancer invasion. By comparison, in the case of fibrinogen, another extracellular matrix molecule involved in blood clotting, knowledge of its adhesion site for platelets has led to the development of clotting pharmaceuticals that are already available to the public.

Similar considerations are applicable to our studies on the MMP2 cleavage site. MMP inhibitors are widely considered strong candidates as anti-metastasis drugs (2). In Aim 2, we have shown that a fragment of Ln-5 is cleaved by MMPs. This result was not obviously predictable, and puts us in a position to eventually use the Ln-5 fragment as a basis for the design of MMP inhibitors.

References:

- 1. Koshikawa, N., Giannelli, G., Cirulli, V., Miyazaki, K., and Quaranta, V. Role of cell surface metalloprotease MT1-MMP in epithelial cell migration over laminin-5. J. Cell Biol. *148*:615-624, 2000.
- 2. Quaranta, V. Cell migration through extracellular matrix: membrane-type metalloproteinases make the way. J. Cell Biol. *149*:1167-1170, 2000.

Appendices:

Letter regarding unpublished data.

Figure 1.

Figure 2.